Mutations in LAMB1 Cause Cobblestone Brain Malformation without Muscular or Ocular Abnormalities

Farid Radmanesh,1,2 Ahmet Okay Caglayan,3,12 Jennifer L. Silhavy,1,2 Cahide Yilmaz,4 Vincent Cantagrel,1,2 Tarek Omar,5 Başak Rosti,1,2 Hande Kaymakcalan,6 Stacey Gabriel,7 Mingfeng Li,8 Nenad Šestan,8 Kaya Bilguvar,3 William B. Dobyns,9,10 Maha S. Zaki,11 Murat Gunel,3,* and Joseph G. Gleeson1,2,*

Cobblestone brain malformation (COB) is a neuronal migration disorder characterized by protrusions of neurons beyond the first cortical layer at the pial surface of the brain. It is usually seen in association with dystroglycanopathy types of congenital muscular dystrophies (CMDs) and ocular abnormalities termed muscle-eye-brain disease. Here we report homozygous deleterious mutations in LAMB1, encoding laminin subunit beta-1, in two families with autosomal-recessive COB. Affected individuals displayed a constellation of brain malformations including cortical gyral and white-matter signal abnormalities, severe cerebellar dysplasia, brainstem hypoplasia, and occipital encephalocoele, but they had less apparent ocular or muscular abnormalities than are typically observed in COB. LAMB1 is localized to the pial basement membrane, suggesting that defective connection between radial glial cells and the pial surface mediated by LAMB1 leads to this malformation.

The cerebral cortex is generated through proliferation of radial glial cells residing in the ventricular zone, migration of postmitotic neurons toward the pial surface, and organization of the laminar architecture with six neuronal layers.1 Radial glial cells play a central role in cerebral cortical development, where they act both as the proliferative unit of the cerebral cortex and a scaffold for neurons migrating toward the pial surface.2 The basement membrane/glia limitans (BM/GL) is located immediately below the pia matter and serves as an anchor point for the endfeet of radial glial cells and as a physical barrier to migrating neurons.3 Mutations in genes encoding BM constituents4–6 those encoding cellular receptors that mediate interactions between cells and the extracellular matrix, or those encoding downstream effectors of such interactions7–9 have been shown to disrupt BM integrity and result in disorganized cortical lamination.

Cobblestone brain malformation (COB, originally referred to as lissencephaly type 2) is characterized by cortical dysplasia and aberrant neuronal migration through breaches in the pial BM/GL into the subarachnoid space;10,11 the result is neuronal ectopia that produce an irregular “lumpy-bumpy” appearance.12,13 COB is typically observed in α-dystroglycanopathy types of congenital muscular dystrophies (CMDs), which are associated with various ocular abnormalities, delineating the spectrum of muscle-eye-brain disease (MIM 236670).14 However, it has also been reported as an isolated entity15 or in conjunction with minimal muscular abnormalities,16 reflecting genetic and clinical heterogeneity of the disorder. In fact, genetic studies have failed to identify a mutated gene in individuals with these forme fruste presentations.17,18 Here we report that mutations in LAMB1, encoding laminin subunit beta-1, are associated with COB with variable muscular or ocular abnormalities. Although LAMB1 is expressed in skeletal muscle,19 our subjects did not have elevated creatine phosphokinase, suggesting LAMB1 mutations as a cause for the muscle-sparing form of CMD.

We recruited two consanguineous families from Egypt and Turkey and report on a total of four affected members (Figure 1). In family 520, the first-cousin parents had three children with severely delayed developmental milestones, increased head circumference, and posterior encephalocoele (Figure S1, available online). Symptomatic hydrocephalus had been present in all three siblings since birth and required shunt placement in the second child. Brain magnetic resonance imaging (MRI) revealed COB, evidenced by abnormal cortical gyri and sulci, dramatic white-matter signal abnormalities, and brainstem and cerebellar hypoplasia (Figure 2). Cortical gyration in the anterior forebrain regions was relatively preserved in comparison with that in the posterior regions on axial MRI sections, suggesting a posterior-anterior gradient. There was diffuse abnormal white-matter signal intensity and subcortical band-like gray matter lying approximately 1 cm below the cortex. The cortex was moderately thick...
and was more pronounced in temporal, parietal, and occipital regions. Deep in the inner cortical surface and separated from the cortex by a layer of white matter, contiguous nodules of gray matter left a thin beaded laminar heterotopia (most evident in subject 520-IV-3), the type of irregular heterotopia seen in other CMDs. There was myelination disruption in subcortical "U" fibers in a pattern typical of COB, as well as delayed myelination in the deep and periventricular white matter. There was also hypoplasia of the brainstem, cerebellar hemispheres, and vermis, as often observed in CMDs. The findings were most consistent with COB type A (the most severe form) according to a recent classification, especially given the presence of encephalocele. Considering the classical association of COB and CMDs, we evaluated these subjects for muscular and ocular manifestations but identified no relevant abnormality (Table S1). They had normal creatine phosphokinase (CPK), electromyography,
and nerve conduction studies and no overt signs of amyotrophy or weakness, so muscle biopsy was not pursued. Ophthalmology assessments with indirect fundoscopy showed diffuse minor optic atrophy in the two older siblings but not in the youngest one (520-IV-3).

In order to identify the genetic cause for this condition, we performed whole-exome sequencing on the middle child in the family. This study was performed in accordance with the ethical standards set by our institutional review boards, and we obtained informed consent from each individual participating in this study. We performed segregation analysis for all unique and conserved variants via Sanger sequencing of PCR products that we amplified by using genomic DNA of all subjects and the parents. These studies led to the identification of a homozygous deletion of 14 bp and an insertion of 41 bp (c.3145_3158delins41) in \( LAMB1 \) (laminin subunit beta-1; Refseq accession number NM_002291.1 [MIM 150240]). This deletion and insertion resulted in a frameshift and premature stop (p.Lys1049Profs*7). The insertion was a triplication of a 19 bp fragment of the gene (c.3159_3177[3]), resulting in the deletion of 14 bases and the insertion of 14 bases and suggesting an inverted repeat-mediated mechanism. The mutation was located in exon 22, encoding a laminin epidermal-growth-factor-like domain (EGFLAM) of the protein (Figures 1E–1F; see also Figure S2). Genome-wide linkage analysis with Illumina linkage map IVb suggested that a locus on chromosome 7q22 carried the mutation with a multipoint logarithm of odds (LOD) score of ~2.3 (Figure 1B). This mapping defined an interval of ~35.5 Mb between the markers rs1029847 and rs885993 and included \( LAMB1 \), supporting the result obtained through whole-exome sequencing. Whole-exome sequencing appropriately covered all genes located in the candidate intervals with highest LOD scores, and no other shared unique deleterious variants were encountered by exome sequencing.

The second family, numbered 1257, was ascertained in Turkey. The parents were first cousins and had a child with a phenotype similar to that of the affected individuals in family 520. They came to medical attention during late gestation because of intrauterine hydrocephalus, and the infant ultimately required shunt placement at 8 months of age. She had seizures, severe developmental delay, and a persistently increased head circumference. Brain MRI revealed COB, band-like heterotopia with posterior predominance, and hydrocephalus (Figure 2), very similar to the appearance of brain MRIs in family 520. Whole-exome sequencing identified a homozygous variant at the first nucleotide of the splice donor site in intron 16 prior to exon 17 of \( LAMB1 \) (c.2110+1G>T). This variant segregated in the family in a way consistent with a recessive mode of inheritance (Figure S3). The mutation is predicted to lead to failure of splicing from exon 16 to exon 17 and to result in a frameshift (p.Ser703fs*62). The variant occurred within a block of homozygosity between chromosome 7 base positions 107575806 and 108524461 (Figure 1D). All genes known to be mutated in CMD were well covered in these subjects, and no deleterious variant was found in any of these genes. Neither of the two mutations was observed in the dbSNP or 1000 Genomes databases or in our cohort of 100 Egyptian and 100 Turkish ethnically matched control subjects, and both were absent from our in-house whole-exome database of more than 2,000 subjects with neurodevelopmental conditions.

Laminins are the major secreted glycoproteins found in the basal lamina, where they influence cell proliferation, differentiation, migration, and adhesion. They are cross-shaped heterotrimeric proteins containing an alpha-, beta-, and gamma-chain from the \( LAMA \), \( LAMB \), and \( LAMC \) families, respectively. These are constituted by
five, four, and three genes of each type, respectively, and contribute to at least 15 different combinations in mammals. Individual laminin subunits demonstrate specific spatial and temporal expression patterns. Laminin subunit beta-1, encoded by \textit{LAMB1}, is present in six of the 15 laminins and is one of the earliest laminin subunits expressed during mammalian development at various sites, including the neuroectoderm. In the nervous system, laminins are thought to be involved in several processes, including neurite outgrowth, neural survival, and synapse formation and function. However, in vivo study of specific laminin subunits in mammalian brain development has been hampered by early lethality of laminin-mutant mice.

We compared the expression of \textit{Lamb1} with the expression of each of the other known \textit{Lamb} genes. We profiled \textit{Lamb1} expression in murine tissue from embryonic day 10 (E10) through postnatal day 15 (P15) by using RT-PCR and compared \textit{Lamb1} transcripts with each of the other known laminin transcripts (Figure 3A). \textit{Lamb1} showed relatively strong and ubiquitous expression in the whole E10 embryo, in E14, E16, P0, and F15 forebrain, and in P15 cerebellum and muscle. Expression in the eye globe was relatively lower than that in other tissues, but expression in embryonic meninges was strong. Mutation of \textit{LAMB3} causes a specific skin defect known as epidermolysis bullosa (MIM 226650), whereas mutations of \textit{LAMB2} predominantly cause a kidney and eye disease known as Pierson syndrome (MIM 609049), which can include intellectual disabilities. \textit{Lamb1} was similar in tissue distribution to \textit{Lamb2}, whereas \textit{Lamb3} was relatively low in each tissue tested (skin was not tested).

Comparing expression of \textit{Lamb1} with the \textit{Lama} and \textit{Lamc} families shows quite similar and broad distributions. \textit{LAMA2} mutations in humans cause Merosin-deficient CMD (MIM 607855), which can include COB-like brain malformations, and \textit{Lama2} showed expression similar to that of \textit{Lamb1}. In addition, we have demonstrated that recessive mutations in \textit{LAMC3}, encoding laminin gamma-3, cause a highly specific occipital pachygyria (MIM 614115) phenotype characterized by cortical thickening and reduced gyration of the occipital lobe, and \textit{Lamc3} shows widespread distribution similar to that of \textit{Lamb1}. In fact, \textit{Lamb2/Lamc3} compound-null mice exhibit severe disruptions in pial BM and cortical ectopias, features that resemble human cobblestone malformation.

Immunostaining of mouse brain at postnatal day 7 with \textit{LAMB1} antibodies demonstrated high levels in the cerebellar BM, consistent with the severe cerebellar phenotype we observed in our subjects (Figure 3B). Zebrafish \textit{lamb1}
mutants have a disintegrated retinal inner limiting membrane and ectopias that protrude into the interstitial space between the retina and the lens.25 These features are consistent with the cortical and cerebellar defects observed in these individuals but suggest that the role for LAMB1 in the eye is not recapitulated in humans.

We next performed weighted gene coexpression analysis by using the human brain transcriptome,33 which we visualized with Cytoscape to explore other genes highly correlated with LAMB1 and genes causing similar brain malformations; such genes include LAMC3, ZIC1, ZIC2, and FLNA (Figure 3C). Zic1 and Zic2 are zinc-finger transcription factors that are critical for proliferation and maintenance of meningeal fibroblasts, and mutant mice show cortical lamination defects resembling COB.34 In addition, heterozygous deletions of ZIC1 and ZIC4 in humans are associated with Dandy-Walker malformation (MIM 220200),35 a severe cerebellar developmental defect. We found that LAMB1 expression was correlated with expression of ZIC1, ZIC2, and genes encoding collagens and other laminins, suggesting a genetic network regulating BM function. Interestingly, COL3A1 was identified at an intersection of LAMB1 and LAMC3. COL3A1 (MIM 606854) encodes a ligand for the receptor GPR56, and mutations in either gene lead to COB-like brain defects in mammals.6,36 Included in this network is NID2, encoding a known component of the BM,37 and COL1A1, which is mutated in Ehlers-Danlos syndrome (MIM 130060) but also produces neocortical dyslaminaton in mice when mutated.38 This correlated expression map suggests a gene network regulating interactions between radial glial cells and the BM.

COB and subcortical band heterotopia (SBH) are categorized as distinct abnormalities because of the shared genetic etiology of SBH with classic lissencephaly (type 1 lissencephaly, MIM [607432]). More recently, a study on the role of RhoA in the developing cerebral cortex has blurred this distinction.39 RhoA is a small GTPase that mediates the linkage between cellular receptors of BM and the actin cytoskeleton. RhoA deletion from radial glial cells specifically causes failure of the cells to span the entire cortical thickness and results in COB and SBH in mice. Similarly, deletion of focal adhesion kinase, a downstream effector of RhoA, from glial cells and meningeal fibroblasts results in abnormalities characteristic for COB.7 These findings suggest that inadequate migration and migration past the pial surface could occur as a result of the same underlying genetic defect. The fact that our subjects share these similar features, including features of COB as well as SBH, supports this model.

All three affected individuals in family S20 had occipital encephalocoele (OE), but the etiology of OE associated with COB is unclear. The major components of the pial BM are produced by meningeal cells,40 and the dura mater contributes essential ossification signals to the overlying cephalic mesenchyme. The dura and arachnoid maters are composed of connective tissue, and therefore defects in laminins could theoretically cause abnormalities in ossificatory signals and thereby protrusion of brain tissue through defects in the skull. Encephalocoeles are known to accompany the severe forms of COB, but the mechanisms are unknown.41,42 Consistent with this, mutation in COL18A1 in humans leads to Knobloch syndrome (MIM 267750), a recessive disorder that includes OE.43 COL18A1 appears adjacent to LAMB1 and LAMC3 on our coexpression map. Because laminins are known to associate with collagens, we hypothesize LAMB1 might associate with COL18A1 at the cephalic mesenchyme.

In summary, we describe loss-of-function mutations in LAMB1 as a cause for COB in the absence of overt muscle or eye abnormalities. Aligning our data with the published data suggests a function for the encoded LAMB1 at the BM, where it mediates both the integrity of the GL and the attachment of radial glial endfeet. As radial glia detach, the scaffolding mediating neuronal migration disintegrates, leading to subcortical heterotopia. At the same time, neurons can migrate past the GL, producing a cobblestone cortex. Further work will be required to determine the exact subunit constituents and genetic requirements of the basal lamina during brain development.

Supplemental Data
Supplemental Data include three figures and one table and can be found with this article online at http://www.cell.com/AJHG/.

Acknowledgments
We are grateful to the families for their participation in the study. We thank the Broad Institute (supported by the US National Human Genome Research Institute; grant U54HG003067 to E. Lander), the Yale Center for Mendelian Disorders; (grant U54HG006504 to R. Lifton, M. Gunel, M. Gerstein, and S. Mane) for sequencing support and analysis, J. Santini at the UCSD Microscopy Core for imaging (P30NS047101), the Yale Biomedical High Performance Computing Center for data analysis and storage, the Yale Program on Neurogenetics, the Yale Center for Human Genetics and Genomics, and Carsten G. Bönnemann (National Institute of Neurological Disorders and Stroke) for discussions. This work was supported by US National Institutes of Health (NIH) grant RC2NS070477 and the Gregory M. Kiez and Mehmet Kutman Foundation (M.G.), NIH grants U01MH081896 (to N.S.), R01NS048453, R01NS052455, and P01HD070494, the Simons Foundation Autism Research Initiative, and the Howard Hughes Medical Institute (to J.G.G.).

Received: October 11, 2012
Revised: November 5, 2012
Accepted: February 8, 2013
Published: March 7, 2013

Web Resources
The URLs for data presented herein are as follows:
Cytoscape, www.cytoscape.org
Human Brain Transcriptome database, http://www.humanbraintranscriptome.org
Accession Numbers

Exome data were deposited at dbGaP under accession number phs000288.v1.p1.

References


Additional references are provided in the document.


Supplemental Data

Mutations in \textit{LAMB1} Cause Cobblestone Brain Malformation without Muscular or Ocular Abnormalities


\textbf{Figure S1. Affected individuals in family 520.} (A-C) Facial photograph of three siblings. (D-F) Photograph of occipital regions depicting encephalocele or scars of encephalocele surgical repair.

\textbf{Figure S2.} \textit{LAMB1} mutation in family 520. Deletion of 14 bases and insertion of 41 bases identified by whole-exome sequencing and confirmed by Sanger sequencing. The sequence chromatogram of a control subject (upper panels), heterozygous parents (middle panels) and patients (lower panels) around the 5` and 3` ends of the INDEL. The arrows indicate the start and end points of the INDEL. The sequence chromatogram of the insertion is depicted at the bottom of the figure.

\textbf{Figure S3.} \textit{LAMB1} mutation in family 1257. The sequence chromatogram of the splice donor site mutation in the affected individual (lower panel) is depicted along with the heterozygous parent and a normal control.
**Table S1.** Summary of clinical features of individuals with homozygous *LAMB1* mutation

<table>
<thead>
<tr>
<th></th>
<th>Family 520</th>
<th>Family 520</th>
<th>Family 520</th>
<th>Family 1257</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>520-IV-1</td>
<td>520-IV-2</td>
<td>520-IV-3</td>
<td>1257-IV-1</td>
</tr>
<tr>
<td><strong>Sex</strong></td>
<td>female</td>
<td>male</td>
<td>male</td>
<td>female</td>
</tr>
<tr>
<td><strong>Age (years)</strong></td>
<td>12</td>
<td>10</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td><strong>Intellectual disability</strong></td>
<td>severe</td>
<td>severe</td>
<td>severe</td>
<td>mild-moderate</td>
</tr>
<tr>
<td><strong>OFC (cm/SD)</strong></td>
<td>55/+1.3</td>
<td>55.8/+2.5</td>
<td>52/+1.5</td>
<td>52/+2</td>
</tr>
<tr>
<td><strong>Motor ability</strong></td>
<td>sitting with support</td>
<td>head control</td>
<td>sitting with support</td>
<td>walking with support</td>
</tr>
<tr>
<td><strong>Hypotonia</strong></td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td><strong>CPK</strong></td>
<td>NL</td>
<td>NL</td>
<td>NL</td>
<td>NL</td>
</tr>
<tr>
<td><strong>EMG/NCV</strong></td>
<td>NL</td>
<td>NL</td>
<td>NL</td>
<td>N/A</td>
</tr>
<tr>
<td><strong>Optic atrophy</strong></td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td><strong>Epileptic seizures</strong></td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><strong>EEG</strong></td>
<td>generalized epileptic activity</td>
<td>bilateral frontotemporoparietal epileptic activity</td>
<td>generalized epileptic activity</td>
<td>frontotemporal epileptic activity</td>
</tr>
<tr>
<td><strong>Hydrocephalus</strong></td>
<td>moderate</td>
<td>severe</td>
<td>moderate</td>
<td>mild</td>
</tr>
<tr>
<td><strong>Encephalocele</strong></td>
<td>occipital</td>
<td>occipital</td>
<td>occipital</td>
<td>-</td>
</tr>
<tr>
<td><strong>Ventriculoperitoneal shunt</strong></td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td><strong>Cobblestone lissencephaly</strong></td>
<td>P&gt;A</td>
<td>P&gt;A</td>
<td>P&gt;A</td>
<td>P&gt;A</td>
</tr>
<tr>
<td><strong>White matter abnormality</strong></td>
<td>diffuse</td>
<td>diffuse</td>
<td>P&gt;A</td>
<td>diffuse</td>
</tr>
<tr>
<td><strong>Corpus callosum</strong></td>
<td>NL</td>
<td>thin</td>
<td>thin</td>
<td>thin</td>
</tr>
<tr>
<td><strong>Cerebellar hemisphere hypoplasia</strong></td>
<td>moderate</td>
<td>severe</td>
<td>moderate</td>
<td>mild</td>
</tr>
<tr>
<td><strong>Cerebellar vermal hypoplasia</strong></td>
<td>severe</td>
<td>severe</td>
<td>severe</td>
<td>moderate</td>
</tr>
<tr>
<td><strong>Brain stem hypoplasia</strong></td>
<td>mild</td>
<td>moderate</td>
<td>moderate</td>
<td>mild</td>
</tr>
<tr>
<td><strong>Cerebellar cysts</strong></td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td><strong>Mutation</strong></td>
<td>c.3145_3158delins41</td>
<td>c.3145_3158delins41</td>
<td>c.3145_3158delins41</td>
<td>c.2110+1G&gt;T</td>
</tr>
</tbody>
</table>

**Abbreviations:** CPK, creatine phosphokinase; EEG, electroencephalogram; EMG, electromyography; N/A, not available; NL, normal; NCV, nerve conduction velocity; OFC, occipitofrontal circumference; P>A, posterior greater than anterior; SD, standard deviation.
Figure S1
Figure S2
Figure S3

c.2110+1G>T

Control

D
G A T

S
T C T G T

Parent

Patient

D
G A T

S
T C T T T